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# Systems for region selective drug delivery in the gastrointestinal tract: biopharmaceutical considerations

Leonid Kagan & Amnon Hoffman

<sup>†</sup>The Hebrew University of Jerusalem, School of Pharmacy, Faculty of Medicine, Department of Pharmaceutics, PO Box 12065, Jerusalem 91120, Israel

Background: The design of a proper region-specific oral drug delivery system has to take into consideration the differences in anatomy, physiology and absorption characteristics that exist between segments within the gastrointestinal (GI) tract, as well as transit kinetics of the dosage form and the site of drug release within the GI tract. Objective: The aim of this review is to examine the various biopharmaceutical aspects of region-specific drug delivery in the GI tract. Methods: This review is mainly focused on pharmacokinetic and pharmacodynamic aspects of regionselective drug delivery with special emphasis on drug absorption pathways (para- and transcellular, absorption and efflux transporters, lymphatic uptake etc.). It includes a discussion of gastroretentive systems, colonic delivery, and lipid-based formulations. The review also addresses targeted therapy of local diseases within the GI tract. Conclusions: The advances in pharmaceutical technology allow for the development of a variety of region-specific drug delivery systems for oral administration to optimize local and systemic therapy.

Keywords: absorption pathways, colonic drug delivery, controlled release, drug absorption transporters, gastroretentive dosage form, peptide delivery

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#### 1. Introduction

The oral route is the most convenient and preferable way to administer drugs. For drugs with high solubility and high permeability (BCS class 1) clinical effectiveness can be achieved by relatively simple immediate or controlled release formulations. However, these formulations can be insufficient for drugs characterized by low systemic absorption following oral administration. The poor bioavailability of pharmacologically active compounds prevents the development of otherwise promising drug candidates and therefore is a major challenge for the pharmaceutical industry. The design of a proper delivery system can modify this unfavorable pharmacokinetic profile and lead to improved bioavailability together with controlled absorption properties.

There are several processes that may occur following drug release from a dosage form in the gastrointestinal (GI) tract, including precipitation, chemical/enzymatic/ bacterial degradation in the lumen, absorption (passive and/or carrier mediated), efflux by P-glycoprotein pump, and metabolism in the intestinal wall. The GI tract is not a uniform structure; it is composed of several regions differing in anatomy, biochemical environment, pH, microbial flora, expression of transporters, and absorption characteristics (Table 1). For drugs that have region-specific absorption properties, transit kinetics of the dosage form (and of the released



Table 1. Parameters of various segments of the gastrointestinal tract [1,3,42,100,113].

| Parameter                  | Stomach                 | Small intestine | Colon               |
|----------------------------|-------------------------|-----------------|---------------------|
| Length (cm)                | 20                      | 350 – 700       | 90 – 150            |
| рН                         | 1.5 – 2 fast, 3 – 6 fed | 6 – 7           | 5.5 – 7             |
| Bacterial count (CFU/ml)   | $10^2 - 10^4$           | $10^3 - 10^4$   | $10^{11} - 10^{12}$ |
| Absolute surface area (m²) | 0.053                   | 200             | 0.35                |
| Transit time (h)           | 0 – 2 fast              | 3 ± 1           | > 20                |

Table 2. Drugs compounded in gastroretentive dosage forms.

| Captopril [114]    | Atenolol [115]         |
|--------------------|------------------------|
| Acyclovir [44]     | Cinnarizine [116]      |
| Cisapride [117]    | Ciprofloxacin [71]     |
| Calcium [118]      | Furosemide [24,70,119] |
| Ketoprofen [120]   | Levodopa [26,121]      |
| Metformin [11,122] | Misoprostol [14]       |
| Sotalol [123]      | Tetracycline [124]     |
| Verapamil [125]    | Diltiazem [126]        |
| Ofloxacin [127]    | Bisphosphonates [128]  |

drug) within the GI tract significantly affect the absorption. The amount of time needed for a dosage form to leave the stomach is highly variable (ranging from several minutes to several hours) [1]. The transit time of the dosage form through the small intestine is relatively constant  $(3 \pm 1 \text{ h})$ and unaffected by food [2,3]. Most of the transit time of a non-disintegrating dosage form in the GI tract is in the large intestine, where it may stay for an extremely long period of time (up to several days) [4]. The pharmacokinetic and pharmacodynamic profiles of the drug may be influenced not only by the physicochemical properties of the drug molecule but also by its delivery site. Successful targeting to an appropriate GI region can lead to enhanced therapy. Local therapy of ailments of the alimentary canal itself (e.g., inflammatory bowel disease [IBD], irritable bowel syndrome, colorectal cancer, and peptic ulcers) is another important reason for targeting a drug to a specific region within the GI tract.

This review focuses mainly on pharmacokinetic and pharmacodynamic aspects of region-selective drug delivery. A brief review of the available technologies and references for more comprehensive technological discussion are also provided. To simplify the discussion we used the term 'conventional' in all cases where the dosage forms were not specially designed to release their payload at a specific region of the GI tract and compared them with targeted drug delivery systems.

## 2. Stomach and upper small intestine

#### 2.1 Technological principles and evaluation

Targeted and controlled delivery of drugs to the upper part of the GI tract can be accomplished by a gastroretentive dosage form (GRDF). Gastric retentivity is achieved by delaying the evacuation of the dosage form from the stomach. The principal challenge in the development of a GRDF is to overcome the normal tendency of the stomach to clear its content (either by continuous propulsive forces in the fed state or by the 'housekeeper waves' occurring every 1 - 2 h in the fasted state). Gastric residence time of the dosage form is variable, and depends on many physiological, pathological and pharmaceutical factors. For example, body posture, age, gender, osmolarity and pH of food are capable of influencing the gastric emptying [1,5].

Several approaches have been employed (with varying degrees of success) to achieve gastric retentivity, including: bioadhesion to gastric mucosa, buoyancy over gastric content, high-density units, administration with pharmacological agents that slow gastric motility, expansion to large dimensions following oral administration, and utilization of an extracorporeal magnet. A detailed appraisal of the various technologies is beyond the scope of this paper and may be found elsewhere [6-10]. Examples of drugs that have been compounded into GRDFs can be found in Table 2.

Administration of the dosage form with high-calorie food is a relatively simple way to attain some degree of gastric retention for a large single non-disintegrating unit [11,12]. These dosage forms are treated by the stomach as indigestible solids and, therefore, their evacuation from the stomach is delayed by repulsive forces in the pyloric region. Hence, a caloric load of the meal (and especially fat content) will dictate the extension of duration of the fed state and thereby gastric retentivity. Providing reproducible gastric retentivity at the fast state is much more complicated and has not yet been achieved [13-15]. The emptying of a multiparticulate dosage form was shown to be less dependent on the presence or absence of food in the stomach [16], and bears the advantage of not being an all-or-nothing system.

When developing region-selective drug delivery systems (such as GRDFs), it is very important to accurately ascertain that the drug release occurs at the desired part of the



GI tract. Therefore, methods to track the transit of the dosage forms in the GI tract are needed. The state-of-the-art technique used for evaluation of the dosage form's performance is gamma scintigraphy [17,18]. This method provides high precision while being relatively safe. A small amount of radioisotopes can be incorporated into the final dosage form without interfering with other components. The direct follow-up on the location of the dosage form provided by gamma scintigraphy can be used for establishing a correlation between GI transit and pharmacokinetic data. Utilization of several isotopes, when one is incorporated into the dosage form and the other is mixed with food, allows for simultaneous determination of the relationship between the dosage form and the food transit time in the GI tract; this may be necessary for the safety evaluation of large gastroretentive devices [19]. Other more accessible methods are X-ray and MRI. MRI is an advantageous imaging technique because it lacks ionizing radiation and therefore allows the researcher to perform experiments in a crossover design [20,21]. One disadvantage is that a relatively large amount of contrast agent may be required to be incorporated into the dosage form to allow detection, which may preclude simultaneous compounding of an active ingredient.

In recent years the development of GRDFs mainly focused on expandable [6,7] and floating techniques [22,23]. Whitehead et al. [15] showed in humans that a multiple unit floating system was significantly more gastroretentive (over 5.5 h) in comparison to non-floating beads in the fed state. A floating tablet of furosemide stayed for 6 h in the stomach of human volunteers [24]. The major limitation of the floating approach is the requirement to maintain a sufficient stomach content to allow an effective separation between the dosage form and the pyloric region [15]. In addition, the gastric retentivity of such dosage forms was shown to be very susceptible to body position [5,25]. We have recently reported that the expanding-unfolding GRDF demonstrated a significant gastric retention (up to 10.5 h in 75% of subjects) when administered with a low-calorie breakfast to healthy volunteers [20]. For this type of GRDF it was demonstrated that the combination of elasticity and geometric properties is a key for ensuring proper gastric retention [26,27]. The major requirements for the expandable devices are an initial small size to be readily swallowed with a rapid size increase in the stomach (to prevent premature emptying), a sufficient final size and strength to withstand stomach contraction, and ultimate biodegradation following the end of drug release. In the development of this type of dosage form safety issues must be taken into consideration, such as possible occlusion of esophagus or pylorus. Moreover, an excessive gastric retention of more than 24 h may result in the accumulation of several dosage units in the stomach. Consequently, the performance of GRDF should be evaluated with various imaging techniques to rule out any possible safety problems.

#### 2.2 Local treatment in the stomach

Continuous delivery of therapeutic agents to the stomach can be used for treatment and prevention of local ailments, such as gastro-esophageal reflux disorder and peptic ulcer. Helicobacter pylori is one of the most prevalent bacterial pathogens and is associated with a variety of disorders (peptic ulcer, gastric cancer, mucosa-associated lymphoma etc.) [28]. Infection by *H. pylori* usually occurs in early childhood, and unless treated, colonization of the gastric mucosa persists for life [28,29]. The treatment of H. pylori associated peptic ulcer is usually based on a combination of several antimicrobial agents with proton pump inhibitors or H<sub>2</sub> blockers; this complex regimen can result in poor compliance. The bacteria reside mainly in the surface layers (mucus and mucosa) of the antral region of the stomach [29,30], and adequately high local antibiotic concentrations are required for successful treatment [31]. The relatively short residence time of conventional dosage forms in the stomach is usually given as the reason that complete eradication of H. pylori is not achieved. It is still unclear whether antimicrobial drugs exert their activity by a local or systemic action following oral delivery [32]. Diffusion from the systemic circulation into the gastric mucosa following intravenous administration was demonstrated in man for several antibiotics, including for example, amoxicillin, metronidazole, and clarithromycin [33]. Despite the controversy, it has been proposed that continuous local delivery could be beneficial for H. pylori therapy. For this reason, several GRDFs that enhance local delivery were evaluated with variable degrees of success. Shah et al. [34] proposed porous chitosan microspheres loaded with amoxicillin and metronidazole. However, they showed a too rapid in vitro release rate. Wang et al. [35] developed positively charged gelatin microspheres of amoxicillin that were mucoadhesive in the isolated rat stomach model. Murata et al. [36] reported that floating alginate gel beads containing chitosan resulted in increased drug concentration in the gastric mucosa compared with metronidazole solution. Rajinikanth and Mishra [37] proposed a floating in situ gelling system of clarithromycin and sucralfate that was found superior to clarithromycin suspension in clearing H. pylori in vivo in Mongolian gerbils. Comprehensive reviews on various drug delivery systems targeted for the treatment of H. pylori can be found elsewhere [29,38].

In general, several points should be taken into consideration when developing a gastroretentive system targeted for the treatment of H. pylori associated ulcers. It seems that expandable large dimension systems may be inadequate and pose some safety problems in cases of mucosal inflammation or ulcer. A multiparticulate system, however, could be advantageous. Moreover, a combination of approaches (like low density and mucoadhesion) can lead to longer retention and improved reproducibility. The major obstacles for mucoadhesive systems are the high turnover rate of gastric mucus and unspecific binding to various intestinal contents.

Still, this type of delivery appears promising for the treatment of H. pylori when close contact with the mucus layer may significantly increase the local antibiotic concentrations. In addition, it is important to note that drugs that significantly elevate gastric pH are usually implemented in H. pylori treatment and utilization of any technology that relies on acidic environment will probably fail to work.

## 2.3 Gastroretentive dosage form and systemic absorption

Following oral administration most drugs have to be absorbed into the blood to produce therapeutic action. The region-specific absorption of certain drugs (sometimes called 'absorption window') may be for various reasons, such as poor solubility at different pH values, poor stability in some GI regions, presence or absence of absorptive or efflux transporters, and presystemic metabolism in the gut wall [39]. The three main absorption mechanisms are transcellular, paracellular, and carrier-mediated transport. For passively transcellularly absorbed drugs no significant variation in the intestinal permeability is expected in the various segments of the alimentary canal. The two other permeation pathways, however, clearly demonstrate regional variability, and these differences are discussed below.

#### 2.3.1 Paracellular absorption route

Small polar molecules are too hydrophilic to penetrate through the cell membrane and therefore are mainly absorbed by the paracellular passive diffusion mechanism. The pore size of epithelial junctions was shown to decrease aboral in the intestine [40-42]. Hydrophilic drugs (e.g., atenolol, acyclovir, metformin) have a higher permeability in the upper GI tract in comparison to lower regions [43]. If such drugs are compounded in a conventional controlled release dosage form, a considerable part of the drug is released in the distal intestine where permeability is limited and thus the bioavailability is impaired. Alternatively, if the drug is slowly released from the GRDF in the upper GI the extent of absorption will be similar to an immediate release formulation. The bioavailability of acyclovir from a magnetic controlled release tablet was reported to be higher when it was positioned in the stomach, with the aid of an external magnet, in comparison to the group without a magnet [44]. We have recently demonstrated in the rat model, that continuous gastric infusion of atenolol results in bioavailability very similar to that of gastric bolus administration (of the same dose), while colonic delivery leads to about a fourfold decrease in the extent of absorption [45]. In summary, the controlled release gastroretentive approach can accomplish prolonged release for hydrophilic drugs without a decrease in bioavailability that occurs with a conventional controlled release formulation.

#### 2.3.2 Carrier-mediated absorption

Drug absorption pathways are not limited to passive permeability, either trans- or paracellular. There are many

carrier-mediated processes controlling the absorption of various compounds in the intestine (amino acids, nucleosides, small peptides, organic cations and anions, phosphates etc.) These transporters are also exploited for absorption of drugs [46-48]. While some of these carriers were shown to be relatively equally distributed throughout the intestine (i.e., organic anion and cation transporters [49]) others clearly exhibited site-specific distribution. For instance, the dipeptide transporter PEPT1, which is involved in the absorption of β-lactam antibiotics and ACE inhibitors, is present almost exclusively in the small intestine [49]. The activity of CNT (nucleoside transporter) was demonstrated to be higher in the proximal intestine in comparison to distal regions [50]. A controlled release gastroretentive mode of administration can considerably alter the pharmacokinetics of drugs absorbed by a carrier-mediated mechanism that is localized in the upper intestine. The impact of this mode of administration depends on the relative capacity of the carriers.

The most prominent effect of GRDFs is expected in drugs that are mainly absorbed from the upper GI tract by a low-capacity carrier. For these drugs a significant increase in bioavailability can be achieved by prolonged delivery to the upper regions of the GI tract in comparison to immediate release formulation. Riboflavin (a water soluble vitamin) is a good model for such drugs [51,52] and is frequently used as a marker compound in the development of GRDFs due to its safety. Several works have demonstrated that prolonged gastric retention can increase the bioavailability of riboflavin by its continuous delivery to the absorbing region at concentrations that are below the level of saturation of the absorption transporters (Figure 1) [20,27,53,54]. Gabapentin, used for treatment of epilepsy and neuropathic pain, is another example. It is absorbed only from the proximal gut by a capacity limited L-amino acid transport system and hence shows dose-dependant bioavailability [55-57]. Development of a controlled-release GRDF led to improvement of dose proportionality [58] and an increase in bioavailability at higher doses [59]. It is of interest that the development of XP13512 (a novel prodrug of gabapentin, engineered to be absorbed by a more equally distributed transporter MCT1) also resulted in enhanced absorption [60].

In contrast to compounds absorbed by saturable carriers, for drugs that are absorbed by high-capacity transporters (localized mainly in the upper intestine) a significant increase in bioavailability following administration by controlled release GRDF should not be expected. We have recently demonstrated that continuous intragastric delivery of valacyclovir (acyclovir prodrug targeted to PEPT1 transporter) allows plasma concentrations to be maintained for a prolonged time in comparison to a high peak followed by rapid elimination after gastric bolus dosing. This concentration profile cannot be attained by a conventional controlled release delivery since the drug is not absorbed from the colon [45]. However, the bioavailability following both gastric bolus and gastric infusion was very similar. In other



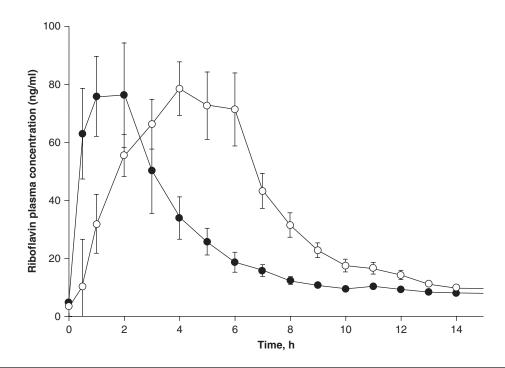


Figure 1. Plasma concentration (mean  $\pm$  SEM) versus time profile of riboflavin. Plasma concentration following administration of the gastroretentive dosage form (GRDF) (O) or the immediate release capsule ( $\bullet$ ) to human volunteers (n = 7). Administration of the GRDF resulted in approximately 1.6-fold increase in bioavailability [20].

words, for these drugs the GRDF approach may provide all the advantages of the controlled release delivery (prevent the high peak concentration, reduce the required doses and improve compliance). But, it should be emphasized that as long as no saturable transport is involved in drug absorption, the GRDF is not expected to increase the bioavailability in comparison to an immediate release formulation.

#### 2.3.3 P-glycoprotein mediated efflux

P-glycoprotein (Pgp) is a well recognized factor in mammalian tumor cell resistance to various cytotoxic drugs and in drug absorption and distribution. It is a plasma membrane-bound ATP-dependent transporter which is encoded in humans by the MDR1 gene. Pgp is found in a wide range of normal tissues and functions to protect the body against potentially toxic xenobiotics. Its localization in the apical membrane of the enterocytes suggests that it may play a major role in limiting the bioavailability of drugs following oral administration. An increasing number of drugs are reported to be substrates of Pgp. Most of the data were obtained from in vitro and ex vivo experiments; however the ability of Pgp to affect drugs' pharmacokinetics was also demonstrated in animal and human studies. It has been suggested that Pgp substrate can benefit if targeted to the upper small intestine [61]. For example, it was demonstrated in one human intestinal perfusion study that the area under the concentration-time curve of talinolol (a known Pgp substrate that is not metabolized by Cyp 3A4 enzyme) decreased with

increasing distance from the teeth [62]. However, the data regarding regional differences in Pgp function in the intestine are rather controversial. It is usually acknowledged that Pgp exhibits a distribution in the intestine that is the opposite of that demonstrated by the Cyp 3A4 enzyme; that is, the lower intestine demonstrates a higher level of Pgp in comparison to the upper intestine. There are several reports that support this statement with respect to the small intestine [63-65]. Different results were obtained when the large intestine was included in the evaluation. Some works showed the highest Pgp level in the colon [66,67], while others reported that the jejunum is the region with maximal Pgp level [68,69]. The variety of methods utilized in the assessment of intestinal Pgp distribution undoubtedly contributes to the variability of the obtained data. In summary, additional research that compares all intestinal parts under the same experimental conditions is required to evaluate the potential advantages of site-specific delivery of Pgp substrates.

#### 2.4 Pharmacodynamic considerations

With the help of gastroretentive technologies it is possible to provide the general advantages of controlled release even to narrow absorption window drugs: reduced frequency of dosing, reduced fluctuation of drug concentration, extended time over critical (effective) concentration, prevention of peak concentration-associated adverse effects etc. In many cases, the pharmacological action of drugs (when given in a bolus mode) on some physiological functions may lead to a

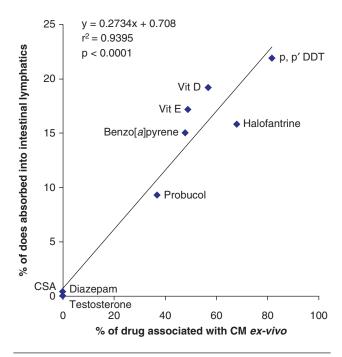


Figure 2. Correlation between the lymphatic availability of tested drugs (% of dose) and degree of association of drugs with isolated chylomicrons in the ex vivo model (% of amount).

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rebound activity of the body that minimizes the therapeutic benefit. Slow input of furosemide (a diuretic drug that has a narrow 'absorption window' in the upper GI tract) from the GRDF has been shown to minimize such counter activity and significantly improve diuretic efficiency of the drug in comparison to an immediate release formulation [70].

Minimization of the local adverse drug effect originating from drug action in the GI tract is also an important issue. A swelling controlled release formulation of ciprofloxacin (designed to deliver its contents over 6 h to the upper GI) demonstrated an antibacterial effectiveness similar to conventional twice daily dosing, while also reducing the incidence of nausea and diarrhea [71]. Retention of the dosage form in the stomach also minimizes the amount of drug that reaches the colon and may reduce adverse effects in it. For example,  $\beta$ -lactam antibiotics are mainly absorbed from the proximal small intestine; the unabsorbed residue can lead to the development of microorganisms' resistance in the large intestine [72].

#### 3. Small intestine

### 3.1 Lipid-based formulations

About 40% of the novel potential drug candidates are highly lipophilic molecules [73] that usually exhibit low oral bioavailability mainly due to low solubility (BCS class 2).

To become orally bioavailable these lipophilic drugs must overcome various barriers. They need to have a sufficient solubility in the aqueous milieu of the GI tract, to penetrate through the unstirred water layer adjacent to the enterocytes, and avoid presystemic metabolism. Dietary fat is absorbed very efficiently in the small intestine, mostly by the lymphatic transport. In order to be absorbed, the dietary lipids need to undergo processing in the GI tract that includes hydrolysis by gastric or pancreatic lipases and emulsification with bile acids and phospholipids (particularly lecithin), which occurs mainly in the upper part of the GI tract. Following oral administration with a high-fat meal or with lipid containing formulation, the lipophilic drug will incorporate into this dietary fat processing mechanism. Administration within lipid-based formulations is a valuable approach to enhance the absorption of lipophilic drugs, and the components of these formulations can enhance bioavailability by several mechanisms.

The lipidic component of the formulation can stimulate biliary secretion of bile salts, phospholipids, and cholesterol, thereby facilitating dissolution. The type of lipidic component has a major influence on the capability of the formulation to enhance absorption. For example, probucol [74] and griseofulvin [75] were better absorbed following administration with medium-chain triglycerides in comparison to long-chain triglycerides. Different lipid-based formulations can be compared using in vitro lipolysis models, which were shown to be predictive for in vivo situations [75,76].

The lipid-based formulation can also improve the absorption of lipophilic drugs by targeting them to intestinal lymphatic uptake [77]. Both the lymphatic system and the circulatory system drain the intestine. The lymph collected from the intestine empties directly into the systemic circulation. Hence, drugs taken up into the lymph bypass the liver and avoid first-pass hepatic metabolism. Each one of the small intestine villi is drained by central lacteal [78]. The lymphatic capillaries of the large intestine are fewer and smaller than the lacteals of the small intestine and located deeper in the intestinal mucosa [79]. The small intestine seems to play a central role in lymphatic uptake. The association of a drug with chylomicrons in the enterocyte is an essential step in the lymphatic absorption pathway. A model that was developed in our laboratory allows this process to be assessed ex vivo and was found to be predictive of the intestinal lymphatic bioavailability potential of lipophilic molecules (Figure 2) [80]. It also makes it possible to estimate the impact of the lymphatic transport on the overall bioavailability [81].

The length of the fatty acid chain of the triglyceride formulation has a great impact on the degree of lymphatic transport. It was demonstrated that long-chain (C<sub>18</sub>) triglycerides lead to a significantly higher lymphatic uptake in comparison to medium-chain (C<sub>8 - 10</sub>) triglycerides of halofantrine [82] or phospholipids-valproic acid conjugate [83]. Development of lipidic prodrugs (by covalent binding



of a fatty acid, a glyceride, or a phospholipid) is a promising approach for enhancement of drug bioavailability by targeting to the lymphatic transport. The rationale is to increase the lipophilicity of the prodrugs to affect their absorption pathway (logP of > 5 appears to be necessary for lymphatic transport) [84]. Several experimental in vivo, in vitro, and in silico methodologies assessment of lymphatic drug transport recently developed [80,81,85-87].

#### 3.2 Ileo-cecal junction

The ileo-cecal junction (ICJ) is located between the small intestine and the colon. Its main physiological function is to regulate chime entry into the large intestine and to prevent the spread of colonic bacterial flora into the small intestine. Although the ICJ may cause a delay in dosage form emptying into the colon of up to several hours [1,4], the size of the dosage form (in a range of 3 - 12 mm in diameter) does not affect the retention time at the ICI [88]. Certain substances, like dietary fat and bile acids, were shown to be able to modulate the ileal brake [89]. This activation of the ileal brake was proposed as a method to prolong the small intestine residence time and thereby to enhance bioavailability [90]. Oleic acid can slow down the small intestine transit rate of non-disintegrating tablets. Dobson et al. [90] evaluated the effect of oleic acid and monoglycerides on atenolol absorption and reported that only in some volunteers did an increase in small intestine transit time lead to an increase in atenolol absorption. It was also emphasized that the prolongation of residence in the upper small intestine has a greater effect on absorption in comparison to residence at the ICJ. Further research is required to evaluate the utilization of the ileal brake on drug absorption.

#### 4. Colon

The colon is significantly different from the small intestine in many aspects. Its primary functions are water and electrolytes absorption and formation and storage fecal material. In contrast to the small intestine there are no villi in colonic epithelium and hence the available area for drug absorption is much smaller. In addition, the amount of fluid is relatively low, especially in the distal colon. Therefore, the colon is frequently omitted from drug absorption considerations. However, the successful performance of controlled release formulations shows that for many drugs the colon demonstrates efficient absorption properties (i.e., metoprolol, isosorbide-5-mononitrate, theophylline, nilsodipine) [42,91]. This is in part due to a prolonged residence time of the dosage form in this region and the low enzymatic degradation capacity in comparison to the small intestine. The colonic region can be effectively used for absorption of highly permeable drugs that are absorbed transcellularly. Delayed release dosage form relying

on colonic absorption may be beneficial for chronotherapy of asthma, ischemic heart disease or arthritis (diseases that exhibit circadian rhythms).

#### 4.1 Colonic delivery technologies for local treatment of IBD

Specific colonic targeted delivery attracted attention mainly due to local therapy in IBD. IBD is a common name several chronic inflammatory conditions of the distal intestine. Conventional drug regimens include aminosalicylates, corticosteroids, antibiotics, and immunosuppressive agents. Since IBD treatments are usually chronic, local delivery of therapeutic drug concentration to the infected regions (in the lower intestine) is preferred to minimize systemic side effects.

Three major strategies are usually implemented separately or in combination for local treatment of IBD and for colonic delivery in general: utilization of a pH drop on entry into the colon that is associated with cecal metabolism of polysaccharides (the mean pH in the distal small intestine and the caecum was reported to be  $7.5 \pm 0.4$  and  $6.4 \pm 0.4$ , respectively [92]); delayed release dosage forms that rely on GI transit time (usually 5 - 6 h); and utilization of metabolic capabilities of colonic bacterial flora to cleave azo and glycosidic bonds [92]. The last approach can be subdivided into two groups: development of prodrugs for specific molecules or design of more universal carrier systems. For example, several azo-bonded modifications of 5-aminosalicylic acid are commercially available to minimize systemic absorption and to maximize local delivery (mesalazine, sulfasalazine, olsalazine, and balsalazide). Alternatively, delivery systems based on various natural polysaccharides have been proposed, including pectin, guar gum, inulin, chitosan, and others [93]. In analogy to azo-bonded compounds there are other redoxsensitive polymers that have been proposed for colonic drug delivery. The mean redox potential in the proximal small intestine, distal small intestine and the colon is  $-67 \pm 90 \text{ mV}$ ,  $-196 \pm 97 \text{ mV}$  and  $-415 \pm 72 \text{ mV}$ , respectively; thus, it can be used as a highly selective mechanism for colonic delivery [94].

approaches, however, have been only partly successful mainly due to a high intra- and interindividual variability. The disease state itself may have a major influence on colonic parameters and thereby affect the in vivo performance of these delivery technologies. For example, it was demonstrated that metabolic activity of the GI tract flora is reduced, colonic pH is lower, and GI transit rate may vary in IBD patients in comparison to the healthy population [95-97]. A combination of several approaches may lead to more reproducible dosage form performance [92,98]. An additional benefit can probably be provided by multiparticulate dosage forms that provide a more uniform GI transit and drug release [4,99].

#### 4.2 Systemic absorption from colon

#### 4.2.1 Peptide and protein delivery

More recently, the colon has attracted much attention as a promising region for delivery of biopharmaceutics [100-102]. Therapeutic proteins have become a considerable part of the arsenal of clinically used drugs. However, the parenteral route of administration continues to be the main delivery route for these agents due to a poor bioavailability following oral administration. The enzymatic degradation in the gut and poor permeability through the intestinal wall are reasons for this low bioavailability. Many works have demonstrated that the magnitude of the proteolytic activity within the colon is significantly lower than in the small intestine and also different from the proteolytic activity of the small intestine and the stomach [100,101,103]. The upper GI tract is a highly aggressive environment for proteins due to a low pH and pepsin secretion in the stomach. The variety of peptidases secreted from the pancreas lead to a rapid protein degradation in the small intestine. Both luminal [104] and brush border membrane [105] protease activity is lower in the colon in comparison to the small intestine and stomach, which makes the colon a preferred region for delivery of therapeutic peptides. Many peptide drugs are hormones, which are very potent, so even small amounts of the absorbed drug can provide a desired therapeutic response. The colon is also considered to be a superior region for the action of absorption enhancers that facilitate drug permeation through the epithelium [106,107].

#### 4.2.2 Absorption of ester prodrugs

Another example of region-specific metabolic activity of the intestine can be illustrated by esterase activity. Colonic delivery may provide certain advantages for delivery of ester prodrugs. Synthesis of ester prodrugs is a valuable approach to enhance the oral drug absorption of highly hydrophilic drugs. The more lipophilic nature of the prodrugs facilitates transcellular permeability. The active moiety is intended to be released following absorption in the systemic circulation. However, the intestinal wall contains a significant amount of esterase activity [108] and this presystemic hydrolysis [109,110] may limit the utility of this approach. Van Gelder et al. [111] evaluated intestinal esterasemediated degradation of p-nitrophenyl acetate and tenofovir disoproxil in several species (rat, man, and pig). They found a significant site-specific difference in esterase activity in all species: duodenum ≥ jejunum > ileum ≥ colon. It was concluded that colonic targeting may be recommended as a potential strategy to reduce esterase-mediated degradation of ester prodrugs.

#### 5. Expert opinion

technological, physicochemical, and engineering knowledge needed to develop drug delivery systems has now reached the stage that it is possible to provide a wide

selection of routes, sites, modes, and rates of administration for a drug. It has previously been established that the development of a drug delivery system should comply with pharmacokinetic considerations, including aspects of absorption, presystemic biotransformation distribution, systemic elimination, and metabolic fate. Pharmacodynamic aspects (including both efficiency and toxicity) that can be influenced by the delivery system should be examined as well.

The drugs that exhibit region-specific absorption properties in the GI tract can benefit from targeted delivery to an appropriate GI segment. For drugs that are highly absorbed but require sustained input function (for example, due to a short half-life), many conventional controlled release technologies can be used. However, for drugs that exhibit poor absorption properties and low bioavailability, more advanced delivery systems are required. These delivery systems can provide a viable solution for low solubility, degradation, and narrow 'absorption window'. The bioavailability, which is an essential issue in oral drug delivery, can be significantly improved in some cases. For drugs that utilize absorption transporters, the proper targeting to the specific region can result in a dramatic change in pharmacokinetic and pharmacodynamic profiles. The pharmaceutical solution of controlled release gastroretentive formulation is still an 'unmet need', and attaining a significant gastric retention is rather difficult. It should be noted that in vitro data (e.g., buoyancy or release rate), as well as retentivity data in animal models do not necessarily reflect the in vivo conditions in humans. Therefore, the development of gastroretentive devices (for human use) must include the assessment of gastric retentivity properties in human volunteers at relatively early stages of development. The prolongation of the GI residence time provided by GRDFs may be utilized to extend the time available for absorption. This may be an advantage not only for narrow 'absorption window' drugs, but for other drugs as well. The development of the gastroretentive approach for oral administration is very promising and further activity in this area of research is expected.

As a consequence of modern drug discovery techniques (i.e., the introduction of combinatorial chemistry together with advances in in vitro screening method) the number of lipophilic drug candidates is constantly increasing. Despite their pharmacological activity, many of these molecules fail to proceed to advanced stages of research and development due to unfavorable properties (e.g., poor water solubility, low bioavailability). The development of a variety of lipidbased formulations provides an opportunity for these drugs to become valuable additions to the arsenal of clinically used drugs. The components of the delivery system can be used to alter not only the absorption process but also the disposition of the lipophilic drugs [112]. It should be taken into consideration that a lipid-based delivery system will be exposed to lipolysis in the GI tract. Thus, there will be a



significant difference between the in vitro release data and the in vivo performance. In general, the absorption of highly lipophilic drugs will take place in the small intestine, in analogy to dietary fat.

The design of appropriate prodrugs for active moieties that have poor pharmacokinetic and pharmacodynamic profiles is another very fascinating approach. It may be used to alter many drug characteristics (i.e., to widen the 'absorption window', to increase solubility or permeability, to target to the lymphatic transport). However, it should be noted that another important advantage of the drug delivery system approach is that a less complicated regulatory approval is required.

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#### **Declaration of interest**

A Hoffman is scientific founder of Intec Pharma Ltd.

#### **Bibliography**

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

- Washington N, Washington C, Wilson CG. Physiological pharmaceutics. Barriers to drug absorption. 2nd edition. New York: Taylor and Francis, Inc.; 2001
- Davis SS, Wilding EA, Wilding IR. Gastrointestinal transit of a matrix tablet formulation: comparison of canine and human data. Int J Pharm 1993;94(1-3):235-8
- Kararli TT. Comparison of the gastrointestinal anatomy, physiology, and biochemistry of humans and commonly used laboratory animals. Biopharm Drug Dispos 1995;16(5):351-80
- Wilding I. Site-specific drug delivery in the gastrointestinal tract. Crit Rev Ther Drug Carrier Syst 2000;17(6):557-620
- A comprehensive review on gastrointestinal physiology and region-specific delivery.
- Timmermans J, Moes AJ. Factors controlling the buoyancy and gastric retention capabilities of floating matrix capsules: new data for reconsidering the controversy. J Pharm Sci 1994;83(1):18-24
- Klausner EA, Lavy E, Friedman M, Hoffman A. Expandable gastroretentive dosage forms. J Control Release 2003;90(2):143-62
- Streubel A, Siepmann J, Bodmeier R. Gastroretentive drug delivery systems. Exp Opin Drug Deliv 2006;3(2):217-33
- A recent review on technological approaches for gastroretentive systems.
- Moes AJ. Gastroretentive dosage forms. Crit Rev Ther Drug Carrier Syst 1993;10(2):143-95

- Reddy LH, Murthy RS. Floating dosage systems in drug delivery. Crit Rev Ther Drug Carrier Syst 2002;19(6):553-85
- 10. Hou SY, Cowles VE, Berner B. Gastric retentive dosage forms: a review. Crit Rev Ther Drug Carrier Syst 2003;20(6):459-97
- 11. Gusler G, Gorsline J, Levy G, et al. Pharmacokinetics of metformin gastric-retentive tablets in healthy volunteers. J Clin Pharmacol 2001;41(6):655-61
- 12. Gusler G, Hou SYE, Berner B, et al. Single dose pharmacokinetics of gabapentin Gastric Retentive (GRTM) tablets in healthy volunteers. Controlled Release Society 31st Annual Meeting; 12 - 16 June 2004; Honolulu, Hawaii, USA
- Agyilirah GA, Green M, duCret R, Banker GS. Evaluation of the gastric retention properties of a cross-linked polymer coated tablet versus those of a non-disintegrating tablet. Int J Pharm 1991;75(2-3):241-7
- Oth M, Franz M, Timmermans J, Moes A. The bilayer floating capsule: a stomach-directed drug delivery system for misoprostol. Pharm Res 1992;9(3):298-302
- Whitehead L, Fell JT, Collett JH, et al. 15. Floating dosage forms: an in vivo study demonstrating prolonged gastric retention. J Control Release 1998;55(1):3-12
- Davis SS, Hardy JG, Taylor MJ, et al. The effect of food on the gastrointestinal transit of pellets and an osmotic device (Osmet). Int J Pharm 1984;21(3):331-40
- Wilding IR, Coupe AJ, Davis SS. The role of gamma-scintigraphy in oral drug delivery. Adv Drug Deliv Rev 2001;46(1-3):103-24
- Davis SS, Hardy JG, Newman SP, Wilding IR. Gamma-scintigraphy in the evaluation of pharmaceutical dosage forms. Eur J Nucl Med 1992;19(11):971-86
- Paper discussing the implementation of imaging in assessment of drug delivery systems performance.

- 19. Afargan M, Kagan L, Kirmayer D, et al. Gastroretentive Accordion pill: Unaltered food transit and enhanced riboflavin bioavailability. Controlled Release Society 33rd Annual Meeting; 22 - 26 July 2006; Vienna, Austria
- 20. Kagan L, Lapidot N, Afargan M, et al. Gastroretentive Accordion Pill: enhancement of riboflavin bioavailability in humans. J Control Release 2006;113(3):208-15
- 21. Steingoetter A, Weishaupt D, Kunz P, et al. Magnetic resonance imaging for the in vivo evaluation of gastric-retentive tablets. Pharm Res 2003;20(12):2001-7
- Arora S, Ali J, Ahuja A, et al. Floating drug delivery systems: a review. AAPS PharmSciTech 2005;6(3):E372-90
- Singh BN, Kim KH. Floating drug delivery systems: an approach to oral controlled drug delivery via gastric retention. J Control Release 2000;63(3):235-59
- Ozdemir N, Ordu S, Ozkan Y. Studies of floating dosage forms of furosemide: in vitro and in vivo evaluations of bilayer tablet formulations. Drug Dev Ind Pharm 2000:26(8):857-66
- 25. Bennett CE, Hardy JG, Wilson CG. The influence of posture on the gastric emptying of antacids. Int J Pharm 1984;21(3):341-7
- Klausner EA, Lavy E, Barta M, et al. Novel gastroretentive dosage forms: evaluation of gastroretentivity and its effect on levodopa absorption in humans. Pharm Res 2003;20(9):1466-73
- 27. Klausner EA, Lavy E, Stepensky D, et al. Novel gastroretentive dosage forms: evaluation of gastroretentivity and its effect on riboflavin absorption in dogs. Pharm Res 2002;19(10):1516-23
- Noonan B, Alm RA. Novel Intervention strategies for Helicobacter pylori treatment. Curr Drug Targets Infect Disord 2002;2(4):331



#### Systems for region selective drug delivery in the gastrointestinal tract: biopharmaceutical considerations

- 29. Conway BR. Drug delivery strategies for the treatment of Helicobacter pylori infections. Curr Pharm Des 2005;11(6):775-90
- 30. Peterson WL. Helicobacter pylori and peptic ulcer disease. N Engl J Med 1991;324(15):1043-8
- 31. Cooreman MP, Krausgrill P, Hengels KJ. Local gastric and serum amoxicillin concentrations after different oral application forms. Antimicrob Agents Chemother 1993;37(7):1506-9
- 32. Lozniewski A, de Korwin JD, Muhale F, Jehl F. Gastric diffusion of antibiotics used against Helicobacter pylori. Int J Antimicrob Agents 1998;9(3):181-93
- 33. Goddard AF, Jessa MJ, Barrett DA, et al. Effect of omeprazole on the distribution of metronidazole, amoxicillin, and clarithromycin in human gastric juice. Gastroenterology 1996;111(2):358-67
- 34. Shah S, Qaqish R, Patel V, Amiji M. Evaluation of the factors influencing stomach-specific delivery of antibacterial agents for Helicobacter pylori infection. J Pharm Pharmacol 1999;51(6):667-72
- 35. Wang J, Tauchi Y, Deguchi Y, et al. Positively charged gelatin microspheres as gastric mucoadhesive drug delivery system for eradication of H. pylori. Drug Deliv 2000;7(4):237-43
- 36. Murata Y, Sasaki N, Miyamoto E, Kawashima S. Use of floating alginate gel beads for stomach-specific drug delivery. Eur J Pharm Biopharm 2000;50(2):221-6
- 37. Rajinikanth PS, Mishra B. Floating in situ gelling system for stomach site-specific delivery of clarithromycin to eradicate H. pylori. J Control Release 2008;125(1):33-41
- 38. Bardonnet PL, Faivre V, Pugh WJ, et al. Gastroretentive dosage forms: overview and special case of Helicobacter pylori. J Control Release 2006;111(1-2):1-18
- 39. Davis SS. Formulation strategies for absorption windows. Drug Discov Today 2005;10(4):249-57
- 40. Rojanasakul Y, Wang LY, Bhat M, et al. The transport barrier of epithelia a comparative study on membrane permeability and charge selectivity in the rabbit. Pharm Res 1992;9(8):1029-34
- 41. Mummaneni V, Dressman JB. Intestinal uptake of cimetidine and ranitidine in rats. Pharm Res 1994;11(11):1599-604

- 42. Rouge N, Buri P, Doelker E. Drug absorption sites in the gastrointestinal tract and dosage forms for site-specific delivery. Int J Pharm 1996;136(1-2):117-39
- An excellent review on drugs with region-specific absorption properties.
- Ungell AL, Nylander S, Bergstrand S, et al. Membrane transport of drugs in different regions of the intestinal tract of the rat. J Pharm Sci 1998;87(3):360-6
- 44. Groning R, Berntgen M, Georgarakis M. Acyclovir serum concentrations following peroral administration of magnetic depot tablets and the influence of extracorporal magnets to control gastrointestinal transit. Eur J Pharm Biopharm 1998;46(3):285-91
- 45. Kagan L, Hoffman A. Selection of drug candidates for gastroretentive dosage forms: pharmacokinetics following continuous intragastric mode of administration in a rat model. Eur J Pharm Biopharm 2008:69:238-46
- 46. Tamai I, Tsuji A. Carrier-mediated approaches for oral drug delivery. Adv Drug Deliv Rev 1996;20(1):5-32
- Sai Y, Tsuji A. Transporter-mediated drug delivery: recent progress and experimental approaches. Drug Discov Today 2004;9(16):712-20
- Steffansen B, Nielsen CU, Brodin B, et al. Intestinal solute carriers: an overview of trends and strategies for improving oral drug absorption. Eur J Pharm Sci 2004;21(1):3-16
- A comprehensive review on intestinal absorption transporters.
- Englund G, Rorsman F, Ronnblom A, et al. Regional levels of drug transporters along the human intestinal tract: co-expression of ABC and SLC transporters and comparison with Caco-2 cells. Eur J Pharm Sci 2006;29(3-4):269-77
- 50. Ngo LY, Patil SD, Unadkat JD. Ontogenic and longitudinal activity of Na+-nucleoside transporters in the human intestine. Am J Physiol Gastrointest Liver Physiol 2001;280(3):G475-81
- Jusko WJ, Levy G. Absorption, metabolism, and excretion of riboflavin-5'-phosphate in man. J Pharm Sci 1967;56(1):58-62
- 52. Levy G, Jusko WJ. Factors affecting the absorption of riboflavin in man. J Pharm Sci 1966;55(3):285-9
- 53. Ahmed IS, Ayres JW. Bioavailability of riboflavin from a gastric retention

- formulation. Int J Pharm 2007;330(1-2):146-54
- Groning R, Cloer C, Georgarakis M, Muller RS. Compressed collagen sponges as gastroretentive dosage forms: in vitro and in vivo studies. Eur J Pharm Sci 2007;30(1):1-6
- Berry DJ, Beran RG, Plunkeft MJ, et al. The absorption of gabapentin following high dose escalation. Seizure 2003;12(1):28-36
- Gidal BE, DeCerce J, Bockbrader HN, et al. Gabapentin bioavailability: effect of dose and frequency of administration in adult patients with epilepsy. Epilepsy Res 1998;31(2):91-99
- 57. Stewart BH, Kugler AR, Thompson PR, Bockbrader HN. A saturable transport mechanism in the intestinal absorption of gabapentin is the underlying cause of the lack of proportionality between increasing dose and drug levels in plasma. Pharm Res 1993:10(2):276-81
- Cowles VE, Gusler G, Gu R, et al. Dose proportionality of gabapentin gastric retentive extended release tablets in beagle dog [abstract T3201]. AAPS J 2006;8(S2)
- Gabapentin extended release Depomed: gabapentin ER, gabapentin gastric retention, gabapentin GR. Drugs R&D 2007;8(5):317-20
- Cundy KC, Annamalai T, Bu L, et al. XP13512 [(+/-)-1-([(alphaisobutanoyloxyethoxy)carbonyl] aminomethyl)-1-cyclohexane acetic acid], a novel gabapentin prodrug: II. Improved oral bioavailability, dose proportionality, and colonic absorption compared with gabapentin in rats and monkeys. J Pharmacol Exp Ther 2004;311(1):324-33
- 61. Friend DR. Drug delivery to the small intestine. Curr Gastroenterol Rep 2004;6(5):371-6
- Gramatte T, Oertel R, Terhaag B, Kirch W. Direct demonstration of small intestinal secretion and site-dependent absorption of the beta-blocker talinolol in humans. Clin Pharmacol Ther 1996;59(5):541-9
- 63. Mouly S, Paine MF. P-glycoprotein increases from proximal to distal regions of human small intestine. Pharm Res 2003;20(10):1595-9
- Yumoto R, Murakami T, Nakamoto Y, et al. Transport of rhodamine 123, a P-glycoprotein substrate, across rat intestine and Caco-2 cell monolayers in the presence of cytochrome P-450 3A-related



- compounds. J Pharmacol Exp Ther 1999;289(1):149-55
- Tian R, Koyabu N, Takanaga H, et al. Effects of grapefruit juice and orange juice on the intestinal efflux of P-glycoprotein substrates. Pharm Res 2002;19(6):802-9
- Fojo AT, Ueda K, Slamon DJ, et al. Expression of a Multidrug-resistance gene in human tumors and tissues. Proc Natl Acad Sci USA 1987;84(1):265-9
- 67. Fricker G, Drewe J, Huwyler J, et al. Relevance of p-glycoprotein for the enteral absorption of cyclosporin A: in vitro in vivo correlation. Br J Pharmacol 1996;118(7):1841-7
- Berggren S, Gall C, Wollnitz N, et al. Gene and protein expression of P-glycoprotein, MRP1, MRP2, and CYP3A4 in the small and large human intestine. Mol Pharm 2007;4(2):252-7
- Nakayama A, Saitoh H, Oda M, et al. Region-dependent disappearance of vinblastine in rat small intestine and characterization of its P-glycoprotein-mediated efflux system. Eur J Pharm Sci 2000;11(4):317-24
- 70. Klausner EA, Lavy E, Stepensky D, et al. Furosemide pharmacokinetics and pharmacodynamics following gastroretentive dosage form administration to healthy volunteers. J Clin Pharmacol 2003;43(7):711-20
- Washington CB, Hou SYE, Campanella C, et al. Pharmacokinetics and pharmacodynamics of a novel extended-release ciprofloxacin in healthy volunteers. J Clin Pharmacol 2005;45(11):1236-44
- 72. Hoffman A, Horwitz E, Hess S, et al. Implications on emergence of antimicrobial resistance as a critical aspect in the design of oral sustained release delivery systems of antimicrobials. Pharm Res 2007 [Epub ahead of print]
- 73. Lipinski CA, Lombardo F, Dominy BW, Feeney PJ. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. Adv Drug Deliv Rev 2001;46(1-3):3-26
- 74. Palin KJ, Wilson CG. The effect of different oils on the absorption of probucol in the rat. J Pharm Pharmacol 1984;36(9):641-3
- Dahan A, Hoffman A. The effect of different lipid based formulations on the

- oral absorption of lipophilic drugs: the ability of in vitro lipolysis and consecutive ex vivo intestinal permeability data to predict in vivo bioavailability in rats. Eur J Pharm Biopharm 2007;67(1):96-105
- 76. Fatouros DG, Mullertz A. In vitro lipid digestion models in design of drug delivery systems for enhancing oral bioavailability. Expert Opin Drug Metab Toxicol 2008;4(1):65-76
- 77. Porter CJH, Charman WN. Intestinal lymphatic drug transport: an update. Adv Drug Deliv Rev 2001;50(1-2):61-80
- O'Driscoll CM. Lipid-based formulations for intestinal lymphatic delivery. Eur J Pharm Sci 2002;15(5):405-15
- Porter CJ. Drug delivery to the lymphatic system. Crit Rev Ther Drug Carrier Syst 1997;14(4):333-93
- Gershkovich P, Hoffman A. Uptake of lipophilic drugs by plasma derived isolated chylomicrons: linear correlation with intestinal lymphatic bioavailability. Eur J Pharm Sci 2005;26(5):394-404
- Gershkovich P, Qadri B, Yacovan A, et al. Different impacts of intestinal lymphatic transport on the oral bioavailability of structurally similar synthetic lipophilic cannabinoids: dexanabinol and PRS-211,220. Eur J Pharm Sci 2007;31(5):298-305
- Caliph SM, Charman WN, Porter CJH. Effect of short-, medium-, and long-chain fatty acid-based vehicles on the absolute oral bioavailability and intestinal lymphatic transport of halofantrine and assessment of mass balance in lymph-cannulated and non-cannulated rats. J Pharm Sci 2000;89(8):1073-84
- Dahan A, Duvdevani R, Shapiro I, et al. The oral absorption of phospholipid prodrugs: in vivo and in vitro mechanistic investigation of trafficking of a lecithin-valproic acid conjugate following oral administration. J Control Release 2008;126(1):1-9
- Dahan A, Hoffman A. Enhanced gastrointestinal absorption of lipophilic drugs. In: Touitou E, Barry BW, editors, Enhancment in drug delivery: CRC Press; 2007. p. 111-31
- Dahan A, Hoffman A. Evaluation of a chylomicron flow blocking approach to investigate the intestinal lymphatic transport of lipophilic drugs. Eur J Pharm Sci 2005;24(4):381-8

- 86. Dahan A, Mendelman A, Amsili S, et al. The effect of general anesthesia on the intestinal lymphatic transport of lipophilic drugs: comparison between anesthetized and freely moving conscious rat models. Eur J Pharm Sci 2007;32(4-5):367-74
- Gershkovich P, Elgart A, Hoffman A. Prediction of lymphatic absorption transport: in-Vivo, ex vivo, in vitro, and in silico models. 6th Annual Meeting of Israeli Chapter of Controlled Release Society; 5 – 6 September 2007; Caesarea, Israel
- 88. Adkin DA, Davis SS, Sparrow RA, Wilding IR. Colonic transit of different sized tablets in healthy subjects. J Control Release 1993;23(2):147-56
- Dobson CL, Davis SS, Chauhan S, et al. The effect of oleic acid on the human ileal brake and its implications for small intestinal transit of tablet formulations. Pharm Res 1999:16(1):92-6
- 90. Dobson CL, Davis SS, Chauhan S, et al. The effect of ileal brake activators on the oral bioavailability of atenolol in man. Int J Pharm 2002;248(1-2):61-70
- 91. Godbillon J, Evard D, Vidon N, et al. Investigation of drug absorption from the gastrointestinal tract of man. 3. Metoprolol in the colon. Br J Clin Pharmacol 1985;19:S113-8
- 92. Basit AW. Advances in colonic drug delivery. Drugs 2005;65(14):1991-2007
- Chourasia MK, Jain SK. Polysaccharides for colon targeted drug delivery. Drug Deliv 2004;11(2):129-48
- 94. Jain A, Gupta Y, Jain SK. Azo chemistry and its potential for colonic delivery. Crit Rev Therap Drug Carrier Syst 2006;23(5):349-400
- 95. Nugent SG, Kumar D, Rampton DS, Evans DF. Intestinal luminal pH in inflammatory bowel disease: possible determinants and implications for therapy with aminosalicylates and other drugs. Gut 2001;48(4):571-7
- 96. Carrette O, Favier C, Mizon C, et al. Bacterial enzymes used for colon-specific drug delivery are decreased in active Crohn's disease. Dig Dis Sci 1995;40(12):2641-6
- 97. Reddy SN, Bazzocchi G, Chan S, et al. Colonic motility and transit in health and ulcerative colitis. Gastroenterology 1991;101(5):1289-97



#### Systems for region selective drug delivery in the gastrointestinal tract: biopharmaceutical considerations

- 98. Rubinstein A. Colonic drug delivery. Drug Discov Today Technol 2005;2(1):33-7
- 99. Asghar LFA, Chandran S. Multiparticulate formulation approach to colon specific drug delivery: current perspectives. J Pharm Pharmaceut Sci 2006;9(3):327-38
- 100. Haupt S, Rubinstein A. The colon as a possible target for orally administered peptide and protein drugs. Crit Rev Therap Drug Carrier Syst 2002;19(6):499-551
- 101. Sinha V, Singh A, Kumar RV, et al. Oral colon-specific drug delivery of protein and peptide drugs. Crit Rev Therap Drug Carrier Syst 2007;24(1):63-92
- 102. Patel M, Shah T, Amin A. Therapeutic opportunities in colon-specific drug-delivery systems. Crit Rev Therap Drug Carrier Syst 2007;24(2):147-202
- A recent review on colonic drug delivery.
- 103. Miura S, Song IS, Morita A, et al. Distribution and biosynthesis of aminopeptidase N and dipeptidyl aminopeptidase-IV in rat small intestine. Biochim Biophys Acta 1983;761(1):66-75
- 104. Ikesue K, Kopeckova P, Kopecek J. Degradation of proteins by guinea pig intestinal enzymes. Int J Pharm 1993;95(1-3):171-9
- 105. Bai JPF. Distribution of brush-border membrane peptidases along the rat intestine. Pharm Res 1994;11(6):897-900
- 106. Ishizawa T, Hayashi M, Awazu S. Enhancement of jejunal and colonic absorption of fosfomycin by promoters in the rat. J Pharm Pharmacol 1987;39(11):892-5
- 107. Sutton SC, Lecluyse EL, Cammack L, Fix JA. Enhanced bioavailability of cefoxitin using palmitoyl L-carnitine. 1. Enhancer activity in different intestinal regions. Pharm Res 1992;9(2):191-4
- 108. Prueksaritanont T, Gorham LM, Hochman JH, et al. Comparative studies of drug-metabolizing enzymes in dog, monkey, and human small intestines, and in Caco-2 cells. Drug Metab Dispos 1996;24(6):634-42
- 109. Inoue M, Morikawa M, Tsuboi M, et al. Comparative study of human intestinal and hepatic esterases as related to enzymatic properties and hydrolyzing activity for ester-type drugs. Jpn J Pharmacol 1980;30(4):529-35

- 110. Inoue M, Morikawa M, Yamada T, et al. Hydrolysis of ester-type drugs by the purified esterase from human intestinal mucosa. Jpn J Pharmacol 1979;29(1):17-25
- 111. Van Gelder J, Shafiee M, De Clercq E, et al. Species-dependent and site-specific intestinal metabolism of ester prodrugs. Int J Pharm 2000;205(1-2):93-100
- 112. Gershkovich P, Hoffman A. Effect of a high-fat meal on absorption and disposition of lipophilic compounds: the importance of degree of association with triglyceride-rich lipoproteins. Eur J Pharm Sci 2007;32(1):24-32
- 113. DeSesso JM, Jacobson CF. Anatomical and physiological parameters affecting gastrointestinal absorption in humans and rats. Food Chem Toxicol 2001;39(3):209-28
- 114. Nur AO, Zhang JS. Captopril floating and/or bioadhesive tablets: design and release kinetics. Drug Dev Ind Pharm 2000:26(9):965-9
- 115. Rouge N, Allemann E, Gex-Fabry M, et al. Comparative pharmacokinetic study of a floating multiple-unit capsule, a high-density multiple-unit capsule and an immediate release tablet containing 25 mg atenolol. Pharm Acta Helv 1998;73(2):81-7
- 116. Machida Y, Inouye K, Tokumura T, et al. Preparation and evaluation of intragastric buoyant preparations. Drug Des Deliv 1989;4(2):155-61
- 117. Wei ZP, Yu ZF, Bi DZ. Design and evaluation of a two-layer floating tablet for gastric retention using cisapride as a model drug. Drug Dev Ind Pharm 2001;27(5):469-74
- 118. Li S, Lin S, Chien YW, et al. Statistical optimization of gastric floating system for oral controlled delivery of calcium. AAPS PharmSciTech 2001;2(1):E1
- 119. Menon A, Ritschel WA, Sakr A. Development and evaluation of a monolithic floating dosage form for furosemide. J Pharm Sci 1994;83(2):239-45
- 120. El-Kamel AH, Sokar MS, Al Gamal SS, Naggar VF. Preparation and evaluation of ketoprofen floating oral delivery system. Int J Pharm 2001;220(1-2):13-21
- 121. Erni W, Held K. The hydrodynamically balanced system: a novel principle of

- controlled drug release. Eur Neurol 1987;27(Suppl 1):21-7
- 122. Stepensky D, Friedman M, Srour W, et al. Preclinical evaluation of pharmacokinetic-pharmacodynamic rationale for oral CR metformin formulation. J Control Release 2001;71(1):107-15
- 123. Chueh HR, Zia H, Rhodes CT. Optimization of sotalol floating and bioadhesive extended release tablet formulations. Drug Dev Ind Pharm 1995;21(15):1725-47
- 124. Hejazi R, Amiji M. Stomach-specific anti-H. pylori therapy. II. Gastric residence studies of tetracycline-loaded chitosan microspheres in gerbils. Pharm Dev Technol 2003;8(3):253-62
- 125. Sawicki W. Pharmacokinetics of verapamil and norverapamil from controlled release floating pellets in humans. Eur J Pharm Biopharm 2002;53(1):29-35
- 126. Gambhire MN, Ambade KW, Kurmi SD, et al. Development and in vitro evaluation of an oral floating matrix tablet formulation of diltiazem hydrochloride. AAPS Pharm Sci Tech 2007;8(3):E1-E9
- 127. Chavanpatil M, Jain P, Chaudhari S, et al. Development of sustained release gastroretentive drug delivery system for ofloxacin: in vitro and in vivo evaluation. Int J Pharm 2005;304(1-2):178-84
- 128. Chauhan H, Shimpi S, Mahadik KR, Paradkar A. Preparation and evaluation of floating risedronate sodium-Gelucire (R) 43/01 formulations. Drug Dev Ind Pharm 2005;31(9):851-60

#### Affiliation

Leonid Kagan<sup>1</sup> MSc Clin Pharm & Amnon Hoffman<sup>†2</sup> <sup>†</sup>Author for correspondence <sup>1</sup>PhD student The Hebrew University of Jerusalem, School of Pharmacy, Faculty of Medicine, Department of Pharmaceutics, PO Box 12065, Jerusalem 91120, Israel <sup>2</sup>Associate Professor and Chairman The Hebrew University of Jerusalem, School of Pharmacy, Faculty of Medicine, Department of Pharmaceutics, PO Box 12065, Jerusalem 91120, Israel Tel: +972 2 6757014; Fax: +972 26757246; E-mail: amnonh@ekmd.huji.ac.il

